



PTF



POTENTIAL BIOLOGICAL ACTIVITY OF CROATIAN TRADITIONAL APPLE CULTIVAR BY-PRODUCTS

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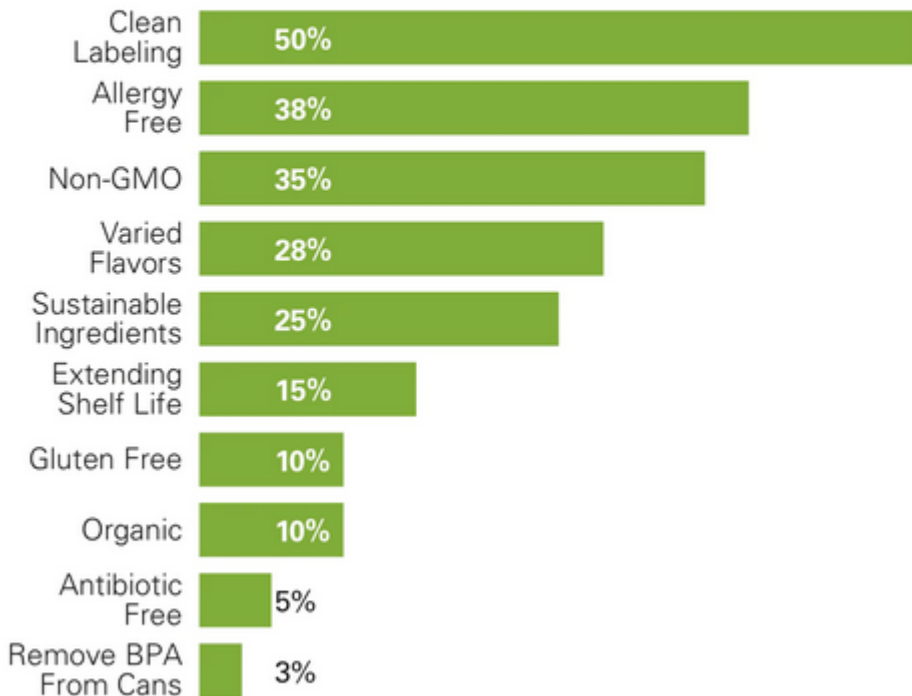
Naziv projekta: Mogućnost iskorištavanja tradicionalnih sorti jabuka za proizvodnju jabuka i soka od jabuka sa smanjenim udjelom patulina

Voditelj projekta:	doc. dr. sc. Ante Lončarić
Izvor financiranja:	Hrvatska zaklada za znanost
Vrijednost projekta:	1.935.745,67 HRK
Trajanje projekta:	1.3.2021. - 28.2.2026.



Clean label

Food companies are impacted by a variety of process changes to fulfill consumer demand and provide a cleaner label:



Source: PMMI Business Intelligence "2019 TRENDS AND ADVANCES IN FOOD PACKAGING AND PROCESSING"

Nowadays, there is an increasing consumer demand for food products with clean labels (Nikmaram et al., 2018). Overall, a clean label product is free of chemically-synthesized and other substances which, despite their origin, are not perceived by consumers as beneficial for their health.



Nikmaram, N., Budaraju, S., Barba, F. J., Lorenzo, J. M., Cox, R. B., Mallikarjunan, K., et al. (2018). Application of plant extracts to improve the shelf-life, nutritional and health-related properties of ready-to-eat meat products. *Meat Science*, 145, 245–255. <https://doi.org/10.1016/j.meatsci.2018.06.031>.

Polyphenols

These nutraceutical compounds can be used in/as;

- food,
- preservatives,
- colourants
- functional foods
- pharmaceutical and cosmetic products
- antimicrobial and fungicides



Antimicrobial polyphenol-rich extracts: Applications and limitations in the food industry



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PLANT MATRICES: SEEDS, FRUIT, PEELS, LEAVES, ROOT, AND WOOD

PLANT EXTRACTS: ETHANOLIC, METHANOLIC, WATER, MICROCAPSULES



FEATURES	APPLICATIONS	CHALLENGES
✓ Antimicrobial Effects	⚙️ Foods	✗ Cost-effectiveness
✓ Antibiofilm Activity	⚙️ Food Packaging	✗ Reproducibility
✓ Consumer Perception	⚙️ Photodynamic Inactivation (PDI)	✗ Sensitivity
✓ No Antimicrobial Resistance	⚙️ Sanitizers	✗ No toxicological Information

Kua, Y. L., Gan, S., Morris, A., & Ng, H. K. (2016). Ethyl lactate as a potential green solvent to extract hydrophilic (polar) and lipophilic (non-polar) phytonutrients simultaneously from fruit and vegetable by-products. *Sustainable Chemistry and Pharmacy*, 4, 21–31. doi:10.1016/j.scp.2016.07.003

Huge volume of food wastes are generated from the processing industry and these low-value food residues are rich in various phytonutrients worth recovering. This approach of valorisation reduces the generation of food wastes and is cost-effective considering the cheap feedstock, reduced waste management expenses and high market value of extracted compounds.

25% of the fresh fruit weight



Water content (66.4-78.2 %)
Sugar content (48.0-62.0 % dw)



'Petrovnjača'



'Paradija'



'Mašanka'



'Slavonska Srčika'

Matrix solid phase dispersion

25 g of dried apple pomace

100 g of sea sand (washed)



Blending

Ratio (g/g)
Dried apple pomace
+
Sea sand (washed)
1:4

Sea sand (washed)
 $m = 20$ g

Glass fibers

Eluent solvents (optima %)

Ethyl lactate	65
Water	35

V_1 (eluent volume) = 500 mL

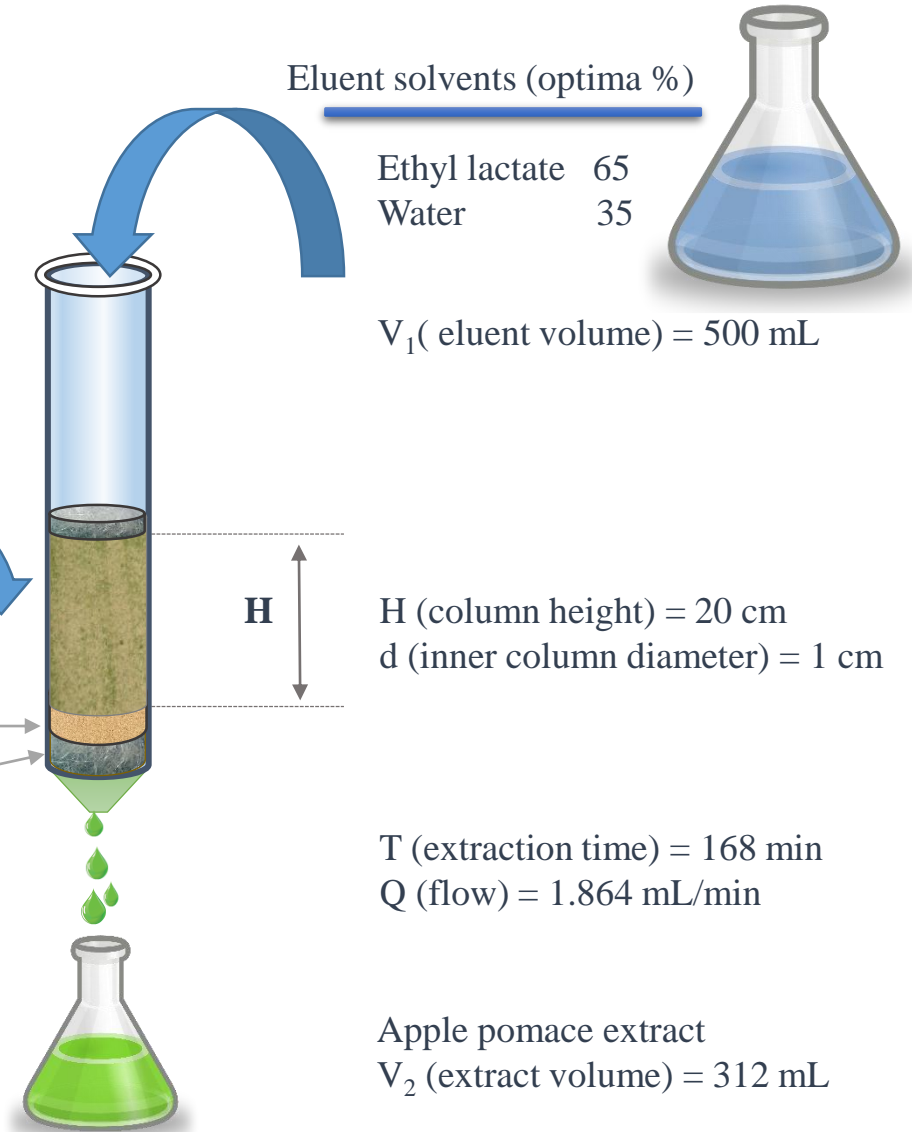
H (column height) = 20 cm
 d (inner column diameter) = 1 cm

T (extraction time) = 168 min

Q (flow) = 1.864 mL/min

Apple pomace extract

V_2 (extract volume) = 312 mL

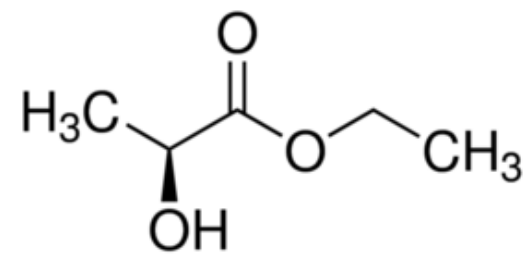


Ethyl lactate



Sustainable Chemistry and Pharmacy

Volume 4, December 2016, Pages 21-31



Volume 500 mL

Solvent: 65% Ethyl lactate

Ratio (g/g); dried apple pomace/sea sand (25g/100g)

Time: 168 min

Extract volume: 312 mL

Ethyl lactate as a potential green solvent to extract hydrophilic (polar) and lipophilic (non-polar) phytonutrients simultaneously from fruit and vegetable by-products

Yin Leng Kua ^a, Suyin Gan ^a, Andrew Morris ^b, Hoon Kiat Ng ^c

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<https://doi.org/10.1016/j.scp.2016.07.003>

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- Ethyl lactate is a green and safe agrochemical solvent.
- Ethyl lactate exerts low toxicity, volatility and flammability.
- Ethyl lactate is able to dissolve in polar and non-polar mediums.
- Research studies have shown its potential in extracting various phytonutrients.
- Ethyl lactate has the potential to extract phytonutrients from food wastes.

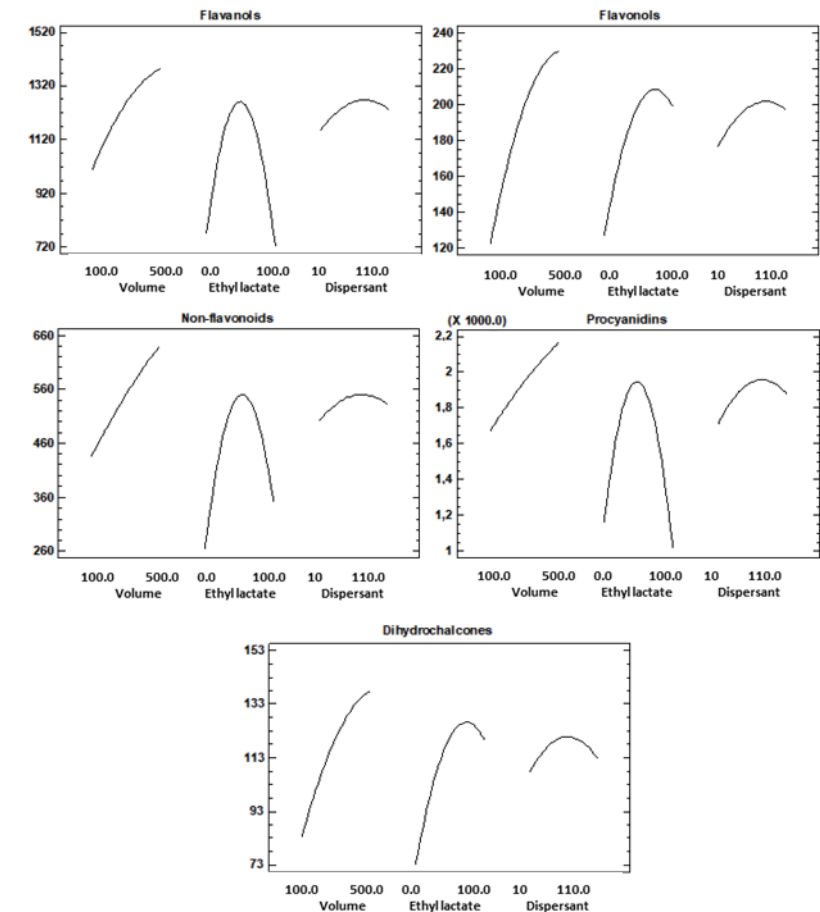


Fig 1. Optimization of extraction of polyphenols from apple pomace

Compounds ($\mu\text{g/mL}$)	Catechin	Epicatechin	Procyanidine B1
'PETROVNJACA'	28.3 \pm 0.26	170.61 \pm 2.17	116.3 \pm 2.9
'WAGNER'	202.84 \pm 5.22	66.71 \pm 0.59	53.23 \pm 0.1
'KLEKER'	131.37 \pm 3.56	143.86 \pm 1.3	187.12 \pm 6.0
'MASANKA'	268.71 \pm 2.27	163.49 \pm 1.57	91.45 \pm 2.1
'ADAMOVKA'	204.07 \pm 2.51	313.54 \pm 1.3	84.76 \pm 0.9
'AMOVKA'	67.55 \pm 0.62	213.88 \pm 2.59	50.68 \pm 1.1
'SRCIKA'	57.04 \pm 0.45	242.53 \pm 1.27	244.51 \pm 4.0
'PARADIJA'	56.24 \pm 2.08	162.34 \pm 1.01	58.44 \pm 0.2

Compounds ($\mu\text{g/mL}$)	Quercetine	Myrecetin	Quercetin 3- β -D-glucoside
'PETROVNJACA'	1167.89 \pm 9.45	35.25 \pm 0.82	19.93 \pm 0.12
'WAGNER'	103.43 \pm 0.98	27.78 \pm 0.16	26.9 \pm 0.07
'KLEKER'	744.35 \pm 18.58	26.94 \pm 0.46	22.29 \pm 0.94
'MASANKA'	880.59 \pm 15.63	47.32 \pm 1.22	35.58 \pm 0.09
'ADAMOVKA'	194.63 \pm 7.33	21.47 \pm 0.26	19.57 \pm 0.52
'AMOVKA'	730.8 \pm 9.05	20.53 \pm 1.29	18.84 \pm 0.36
'SRCIKA'	469.93 \pm 7.13	37.72 \pm 1.93	21.26 \pm 0.36
'PARADIJA'	958.03 \pm 6.3	33.9 \pm 1.36	22.14 \pm 0.79

Compounds ($\mu\text{g/mL}$)	Chlorogenic acid	4-Hydroxycinnamic acid	2-6-dimethoxybenzoic acid
'PETROVNJACA'	1429.22 \pm 16.03	48.64 \pm 1.01	37.38 \pm 0.75
'WAGNER'	639 \pm 1.71	7.45 \pm 0.14	25.69 \pm 0.31
'KLEKER'	1131.27 \pm 16.58	32.7 \pm 0.48	25.36 \pm 0.77
'MASANKA'	1657.98 \pm 14.01	14.97 \pm 0.61	11.7 \pm 0.42
'ADAMOVKA'	900.28 \pm 7.24	64.58 \pm 0.4	36.83 \pm 0.72
'AMOVKA'	1194.39 \pm 21.63	22.29 \pm 0.24	16.65 \pm 0.51
'SRCIKA'	1715.54 \pm 13.38	73.56 \pm 0.76	32.15 \pm 0.28
'PARADIJA'	2383.49 \pm 47.14	28.8 \pm 0.31	33.47 \pm 0.23

Compounds ($\mu\text{g/mL}$)	Phloridzin	Phloretin	Total dihydrochalcones
'PETROVNJACA'	101.58 \pm 1.25	18.22 \pm 0.55	119.8
'WAGNER'	155.11 \pm 1.61	30.14 \pm 0.09	185.2
'KLEKER'	121.58 \pm 2.02	20.06 \pm 1.29	141.6
'MASANKA'	311.97 \pm 1.09	40.36 \pm 1.64	352.3
'ADAMOVKA'	49.26 \pm 1.07	11.93 \pm 0.29	61.2
'AMOVKA'	39.6 \pm 0.91	17.27 \pm 1.87	56.8
'SRCIKA'	161.37 \pm 1.67	20.75 \pm 0.25	182.1
'PARADIJA'	85.23 \pm 0.57	21.98 \pm 0.61	107.2



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Review

Bioactive and functional compounds in apple pomace from juice and cider manufacturing: Potential use in dermal formulations

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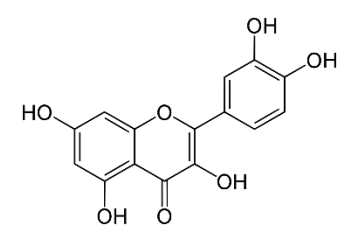
Table 1

Variation in phenolic content in the pomace of different apple cultivars (Ramirez-Ambrosi et al., 2013; Sánchez-Rabaneda et al., 2004).

Compound	Concentration (g/kg DM)	Reference
Phenolic acids		
Protocatechuic acid	0.03–0.26	(Diñeiro García, Valles, & Picinelli Lobo, 2009; Suárez et al., 2010)
Caffeic acid	0.02	(Suárez et al., 2010; Četković et al., 2008)
p-Coumaroylquinic acid	0.09	Andreas Schieber et al. (2003)
p-Coumaric acid	0.02	Andreas Schieber et al. (2003)
Ferulic acid	0.02	Andreas Schieber et al. (2003)
Chlorogenic acid	0.03–0.40	(Diñeiro García et al., 2009; Andreas Schieber et al., 2003; Suárez et al., 2010; Četković et al., 2008)
Caffeic acid	0.28	Lu and Foo (1997)
Flavonoids		
Epicatechin	0.02–0.64	(Diñeiro García et al., 2009; Lu & Foo, 1997; Andreas Schieber et al., 2003; Suárez et al., 2010; Četković et al., 2008)
Catechin	0.12	Andreas Schieber et al. (2003)
Procyanidin B2	0.18–0.45	(Diñeiro García et al., 2009; Andreas Schieber et al., 2003; Suárez et al., 2010)
Quercetin-3-galactoside	0.57–1.61	(Lu & Foo, 1997; Andreas Schieber et al., 2003)
Quercetin-3-glucoside	0.25–0.87	(Lu & Foo, 1997; Andreas Schieber et al., 2003)
Quercetin-3-xyloside	0.09–0.53	(Lu & Foo, 1997; Andreas Schieber et al., 2003)
Quercetin-3-rutinoside	0.06–0.48	(Andreas Schieber et al., 2003; Četković et al., 2008)
Quercetin-3-araboside	0.98	Lu and Foo (1997)
Quercetin-3-rhamnoside	0.05–0.47	(Lu & Foo, 1997; Andreas Schieber et al., 2003)
Hyperin	0.11–0.23	(Diñeiro García et al., 2009; Suárez et al., 2010)
Isoquercitrin + rutin	0.06–0.16	(Diñeiro García et al., 2009; Suárez et al., 2010)
Reynoutrin	0.02–0.08	(Diñeiro García et al., 2009; Suárez et al., 2010)
Avicularin	0.09–0.26	(Diñeiro García et al., 2009; Suárez et al., 2010)
Quercitrin	0.09–0.19	(Diñeiro García et al., 2009; Suárez et al., 2010)
Di-hydrochalcones		
3-Hydroxyphloridzin	0.27	Lu and Foo (1997)
Phloretin-2'-xyloglucoside	0.02–0.40	(Diñeiro García et al., 2009; Lu & Foo, 1997; Andreas Schieber et al., 2003; Suárez et al., 2010)
Phloridzin	0.01–2.00	(Diñeiro García et al., 2009; Lu & Foo, 1997; Andreas Schieber et al., 2003; Suárez et al., 2010; Četković et al., 2008)



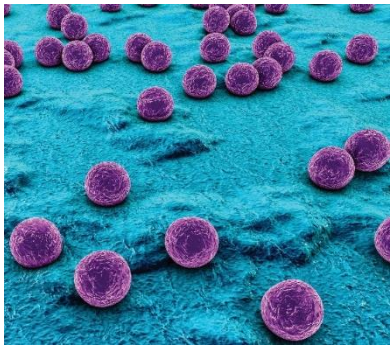
Antimicrobial Activity



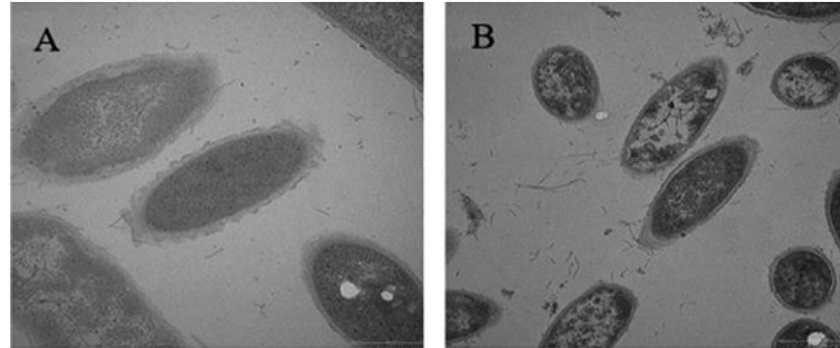
Quercetin



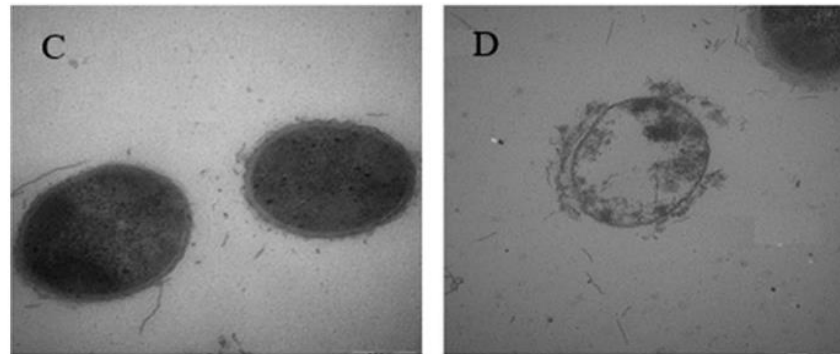
Escherichia coli



Staphylococcus aureus



E. coli (50 MIC)

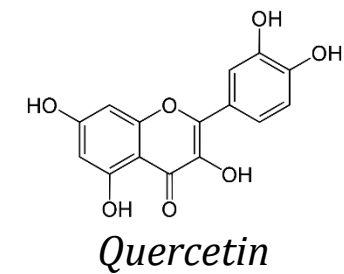


S. aureus (10 MIC)

- **Quercetin** damaged the structure of the bacterial cell wall and cell membrane, leading to increased permeability of these structures.
- The endochylema contents of the cell were released and the activity of ATP was affected.
- Quercetin decreases the synthesis of bacterial proteins, affected the expression of proteins in the cell, and finally resulted in cell lysis and death.

Effects of quercetin on cell wall ultrastructure of *E. coli* and *S. aureus*. (A) *E. coli* without quercetin; (B) *E. coli* with quercetin; (C) *S. aureus* without quercetin; (D) *S. aureus* with quercetin (Wang et al. 2018).

Wang, S.; Yao, J.; Zhou, B.; Yang, J.; Chaudry, M.T.; Wang, M.; Xiao, F.; Li, Y.; Yin, W. Bacteriostatic effect of quercetin as an antibiotic alternative in vivo and its antibacterial mechanism in vitro. *J. Food Prot.* **2018**, 81, 68–78.



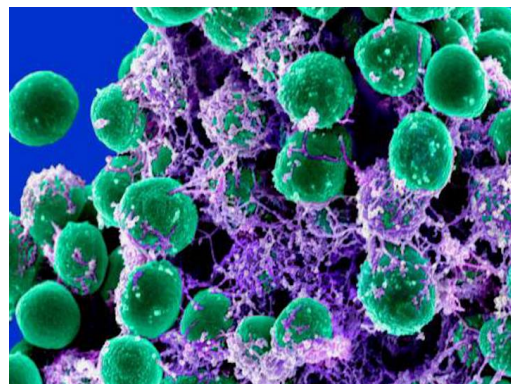
Quercetin exhibits potent bacteriostatic activity against different strains of bacteria, however with more effectiveness against Gram-positive than Gram-negative bacteria.



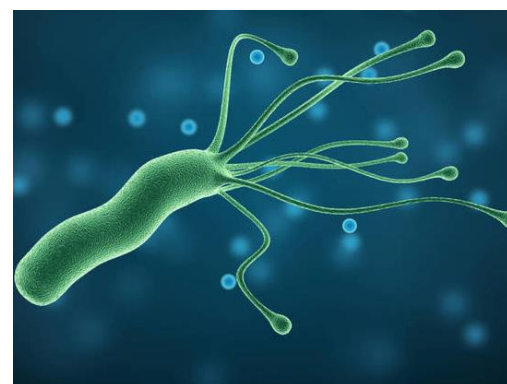
Campylobacter jejuni



Pseudomonas aeruginosa



Staphylococcus epidermidis



Helicobacter pylori



Salmonella enterica

Wang, S.; Yao, J.; Zhou, B.; Yang, J.; Chaudry, M.T.; Wang, M.; Xiao, F.; Li, Y.; Yin, W. Bacteriostatic effect of quercetin as an antibiotic alternative in vivo and its antibacterial mechanism in vitro. *J. Food Prot.* **2018**, *81*, 68–78. [CrossRef]

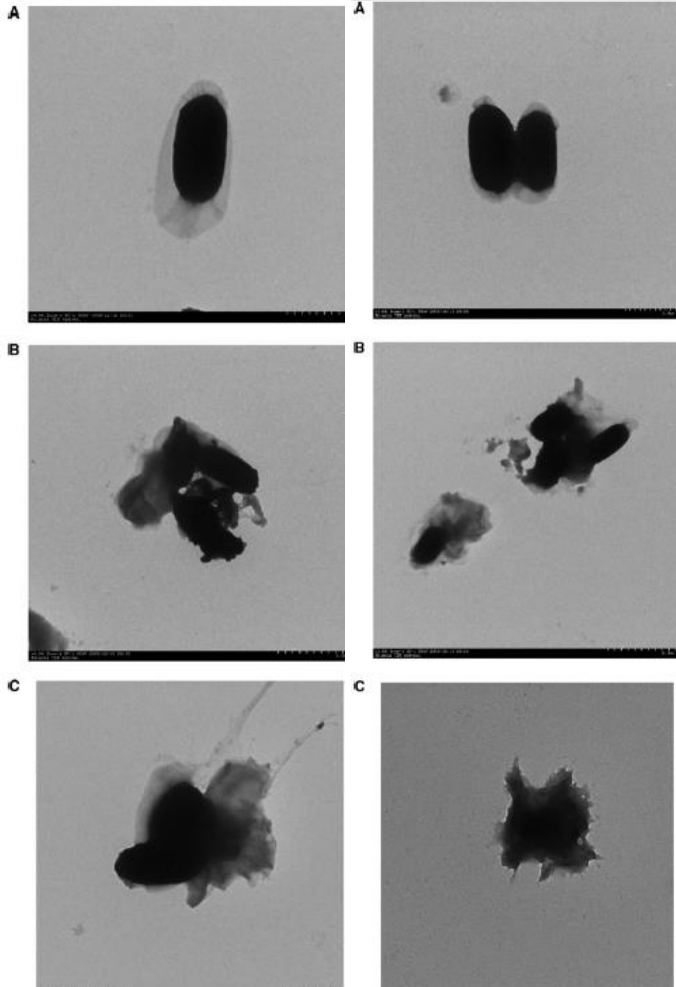
Jaisinghani, R. Antibacterial properties of quercetin. *Microb. Res.* **2017**, *8*. [CrossRef]

Osonga, F.J.; Akgul, A.; Miller, R.M.; Eshun, G.B.; Yazgan, I.; Akgul, A.; Sadik, O.A. Antimicrobial activity of a new class of phosphorylated and modified flavonoids. *ACS. Omega* **2019**, *4*, 12865–12871. [CrossRef]

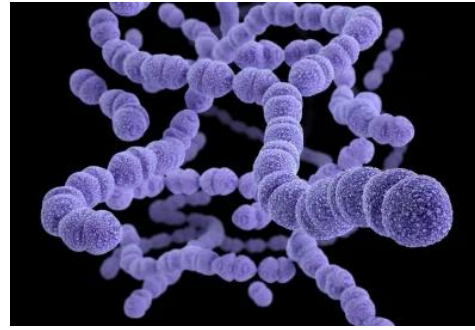
Chlorogenic acid - effectively inhibited the growth of bacterial pathogens;



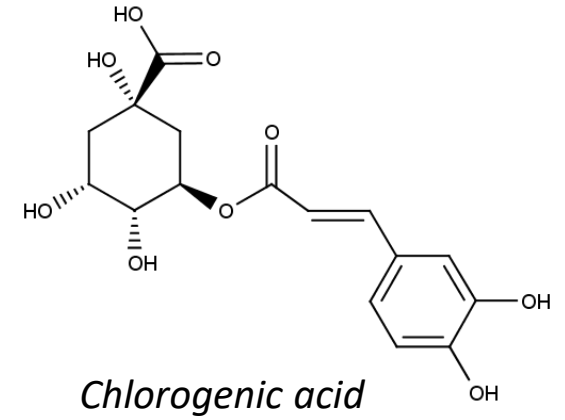
Shigella dysenteriae



Transmission electron micrographs of *Shigella dysenteriae* (left) *Streptococcus pneumoniae* (right) treated by chlorogenic acid for 90 min (B) and 3 h (C), (A) control (Lou et al. 2011).



Streptococcus pneumoniae



Journal of
Food Science

A Publication of
the Institute of Food Technologists

Antibacterial Activity and Mechanism of Action of Chlorogenic Acid

Zaixiang Lou, Hongxin Wang, Song Zhu, Chaoyang Ma, Zhouping Wang

First published: 02 June 2011 | <https://doi.org/10.1111/j.1750-3841.2011.02213.x> | Citations: 248

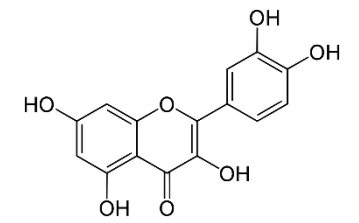
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Mechanism of action - significantly increased the outer and plasma membrane permeability, resulting in the loss of the barrier function, even inducing slight leakage of nucleotide, which led to cell death.

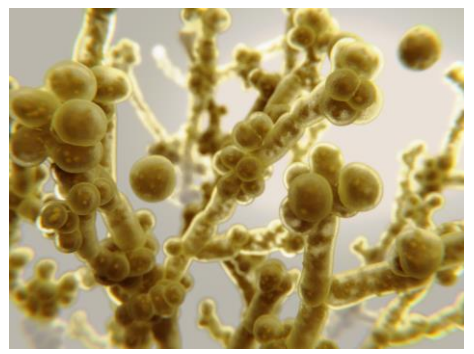
Minimum inhibitory concentration - 20 to 80 µg/mL

Antifungal Activity

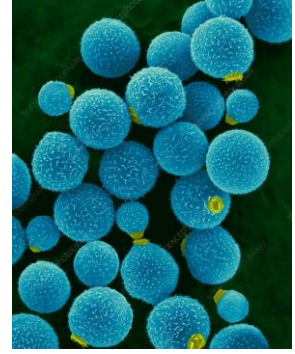


Quercetin

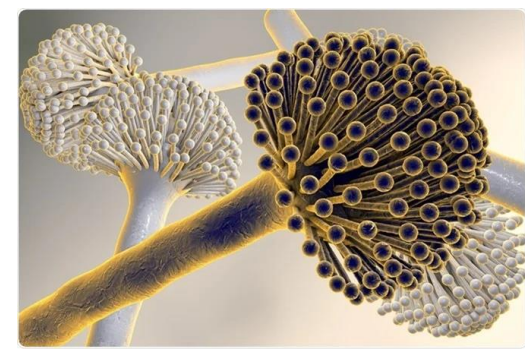
Quercetin revealed the strongest antifungal activities against (Abd-Allah et al. 2015):



Candida albicans

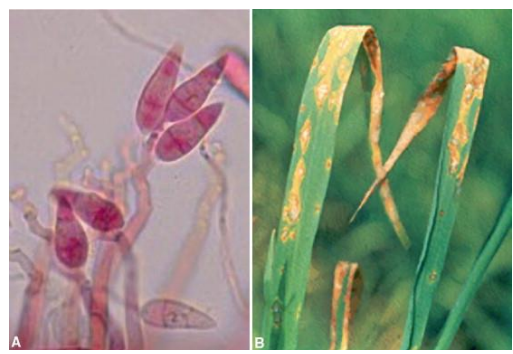


Cryptococcus neoformans



Aspergillus niger

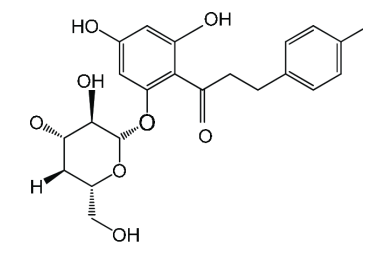
Phloridzin & Phloretin (Shim et al., 2010)



Magnaporthe grisea



Phytophthora infestans

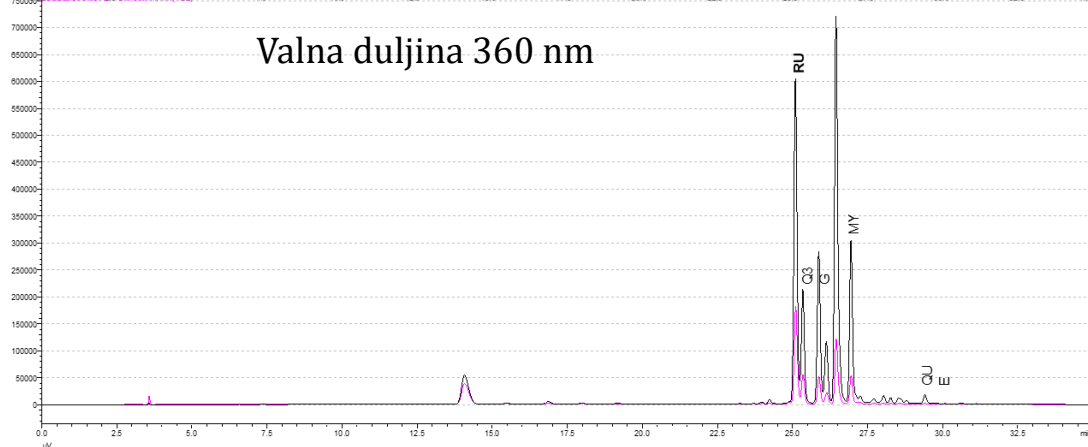
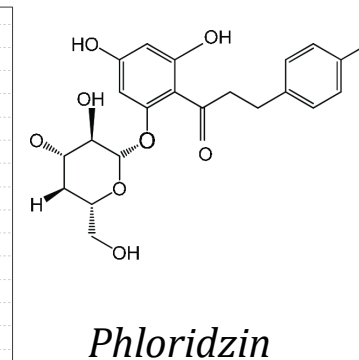
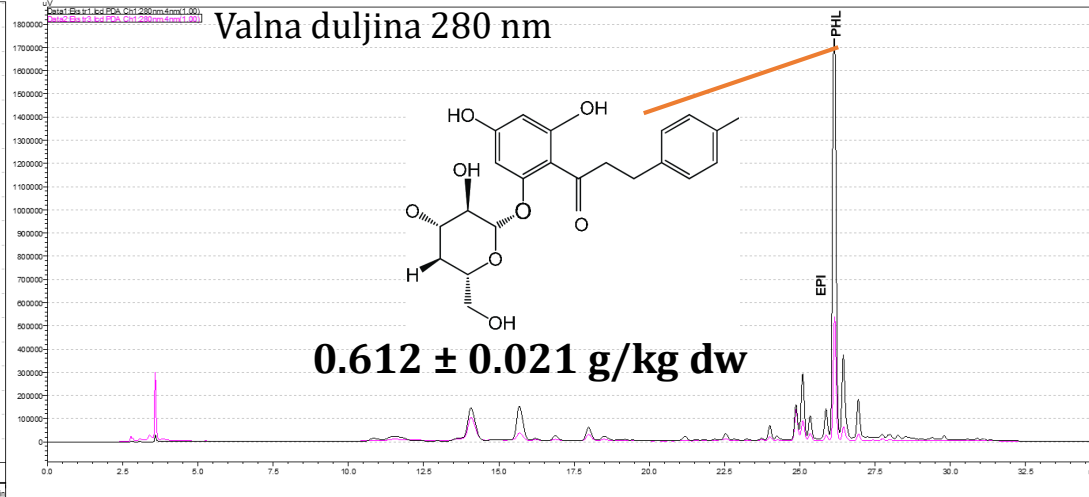
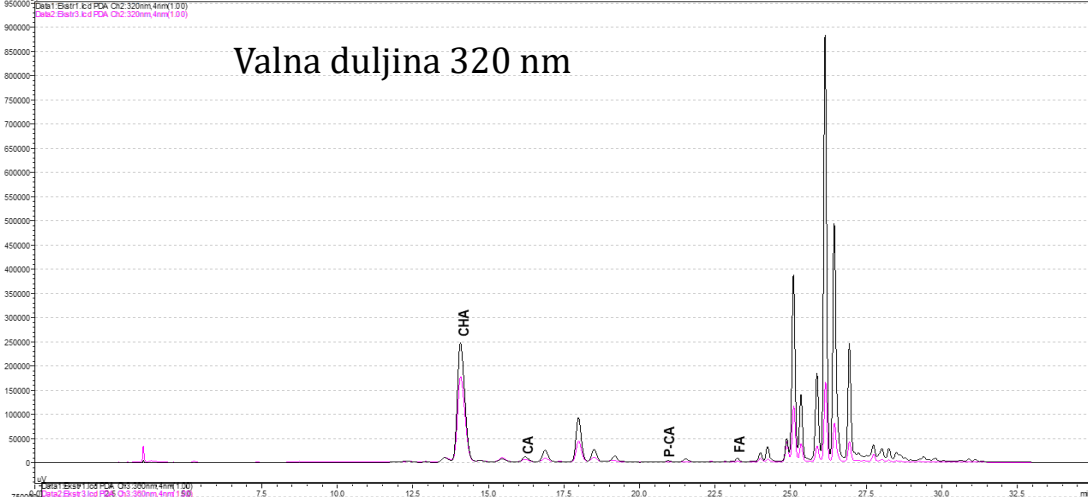


Phloridzin

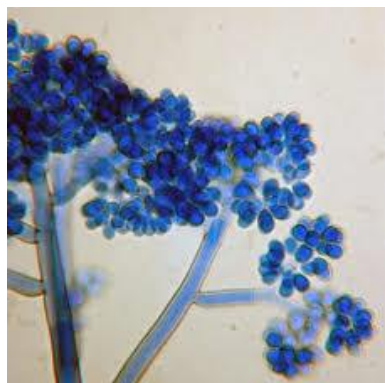
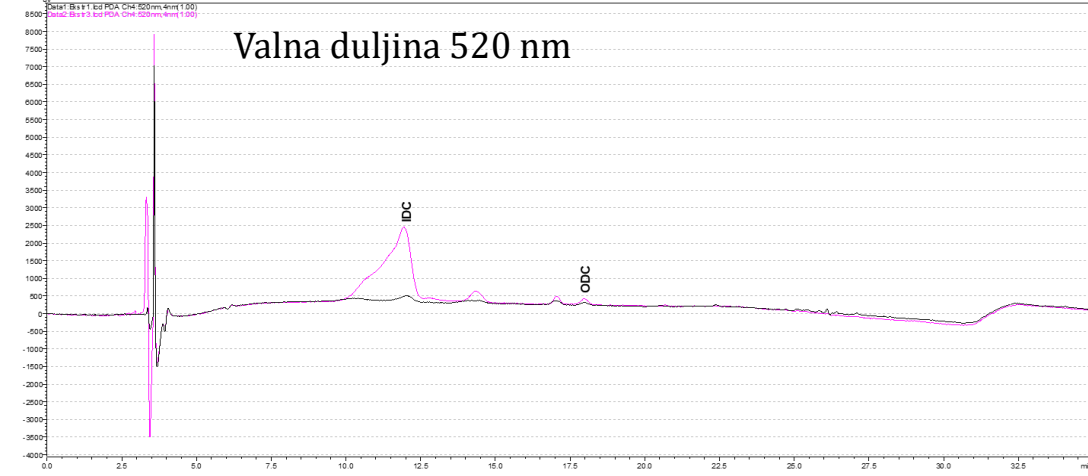
Minimum inhibitory concentration - 67 to 200 µg/mL

Abd-Allah, W.E.; Awad, H.M.; Abdel Mohsen, M.M. HPLC analysis of quercetin and antimicrobial activity of comparative methanol extracts of *Shinus molle* L. *Int. J. Curr. Microbiol. Appl. Sci.* **2015**, 4, 550-558.

Sang-Hee Shim, Su-Jung Jo, Jin-Cheol Kim and Gyung Ja Choi. Control Efficacy of Phloretin Isolated from Apple Fruits Against Several Plant Diseases *Plant Pathol. J.* 2010., 26(3):280-285



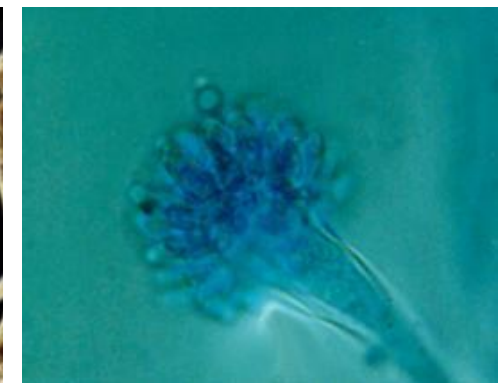
Phloridzin - plays a major role in apple in the resistance to fungal infection. It is metabolized to phloretin and then, to the next oxidation products such as o-quinone, which are fungitoxic (Oleszek et al. 2019).



Botrytis cinerea



Fusarium oxysporum



Neosartorya fischeri

Minimum inhibitory concentration <math>< 500 \mu\text{g/mL}</math>

By-Product	Type of application	Target Microorganisms	References
Apple skin	Chitosan edible film	<i>E. coli</i> , <i>S. enterica</i> and <i>L. monocytogenes</i>	Riaz et al. 2018
Apple skin (powder, extract)	Composite films ASP/CMC	<i>L. monocytogenes</i> , <i>S. aureus</i> , <i>S. enterica</i> and <i>S. flexner</i>	Choi et al. 2017
Apple pomace	Apple phenolic fractions acetone/ethanol	<i>L. monocytogenes</i> , <i>S. aureus</i> , <i>E. coli</i> , <i>S. Typhimurium</i>	Zhang et al. 2016
Apple pomace	Phenolic extract ethyl acetate	<i>S. aureus</i> , <i>E. coli</i>	Raphaelli et al., 2019
Apple peel	Phenolic extract Ethanol/water	<i>Lactobacillus acidophilus</i> , <i>Lactobacillus bulgaricus</i> , <i>Lactobacillus plantarum</i> , <i>Lactobacillus rhamnosus</i> , <i>E. coli</i> , <i>Bacillus cereus</i> , <i>Staphylococcus aureus</i>	Fратиanni et al. 2011
Apple pomace	Fraction of phenolic extract	<i>Botrytis cinerea</i> ; <i>Fusarium oxysporum</i> ; <i>Neosartorya fischeri</i>	Oleszek et al. 2019
Apple pomace	Isolated phloretin	<i>Phytophthora capsici</i> ; <i>Alternaria panax</i> ; <i>Magnaporthe grisea</i> ; <i>Sclerotinia sclerotiorum</i> ; <i>Rhizoctonia solani</i>	Shim et al. 2010

Riaz, A.; Lei, S.; Akhtar, H.M.S.; Wan, P.; Chen, D.; Jabbar, S.; Abid, M.; Hashim, M.M.; Zeng, X. Preparation and characterization of chitosan-based antimicrobial active food packaging film incorporated with apple peel polyphenols. *Int. J. Biol. Macromol.* **2018**, *114*, 547–555. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]

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Conclusions

- MSPD method proves to be an inexpensive and simple method for polyphenol extraction, it could allow the separation of individual polyphenol fractions
- Ethyl-Lactate gives extract rich in polyphenols
- Croatian apple cultivars, 'Petrovnjača', 'Mašanka', 'Paradija' contains polyphenols (quercetin, phloridzin, and chlorogenic acid, respectively) with potential antimicrobial and antifungal activity



PTF



**Hvala na pažnji i
lijepi pozdrav iz
Santiaga de
Compostele!**

